Phylogenetic methods for quantitative trait mapping with complex data sets

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OUTLINE

- **•** Introduction
- **•** Motivation
- **•** Proposed Methods
- **•** Simulation Study
- **Conclusions**
- **Future Directions**

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INTRODUCTION

Motivation: Search for Single Nucleotide Polymorphisms (SNPs) and/or external covariates associated with quantitative traits

Goals:

- **Detection of Associated SNPs**
- Localization of Associated SNPs
- **Detection of Associated Covariates**

Aims of Proposed Work:

- Combine ideas from stochastic processes and phylogenetics to simulate genetic and trait data
- Identify SNPs and/or external covariates associated with quantitative traits using a proposed likelihood score statistic

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EXAMPLE: Outbred Mice Study (Zhang et al. 2012)

Mice Data:

- Organisms: 288 outbred male mice
- **Genetic Data: Genome-wide Association Study SNP data**
- Quantitative Trait Data: High-Density Lipoprotein (HDL) level for each mouse

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EXAMPLE: Deer Mice Study (Linnen et al. 2013)

Deer Mice (*Peromyscus maniculatus*) Study:

- Organisms: 91 wild-caught mice from the edge of the Nebraska Sand Hills
- **Genetic Data: SNP data**
- Quantitative Traits: nine quantitative color phenotypes
- Covariates: weight, body length, tail length, ear length, foot length, sex, and pregnancy status Figure 2A, Linnen et al. 2013

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EXAMPLE: Deer Mice Study (Linnen et al. 2013)

- **•** Researchers are interested in identifying regions of the genome contributing to mouse coat color.
- **Previous work has shown that** much of the variation in coat color appears to be controlled by a single gene, Agouti.

Dark Phenotypes

Figure 2A, Linnen et al. 2013

INTRODUCTION: The Data

Phenotypic Data:

- Quantitative trait data
- One observation per individual in the study

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SNP Data:

Can be represented as a collection of binary random variables

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Data Example: (3 diploid individuals)

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INTRODUCTION: The Data

Quantitative Trait Data:

• One observation per individual in the study

SNP Data:

Can be represented as a collection of binary random variables

Covariate Data:

• One observation per individual in the study

Data Example: 3 diploid individuals

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INTRODUCTION: Previous Methods

Regression-based Methods:

- Tend to detect large genetic signals
- Assume observations have means that are related directly to their genotype and covariate value.
- Assume observations are independent

Phylogenetic Methods:

- Use the relationships within each SNP to gain information about the correlation structure among individuals.
- Use this correlation structure to help improve data analysis.
- Assume observations are normally distributed
- Assume observations have means that related directly to their covariate value and their evolutionary history.

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INTRODUCTION: Motivating Example

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INTRODUCTION: The Data

How the Data are Used:

- Use SNP data to learn about evolutionary relationships
- **Use trait data and covariate data** to find connections between the trait and the SNP and/or the covariate

Note:

- Relationships among SNPs exist due to evolution of genetic data.
- Relationships among trait values are imposed by the relationships among SNPs and environmental covariates.

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INTRODUCTION: The Phylogenetic Framework

Figure: SNPs along a Chromosome

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PHYLOGENETIC METHOD: Step 1

At each SNP,

- Estimate or use the underlying phylogenetic tree, Θ.
- Partition the estimated tree into k clusters using the $(k 1)$ earliest edges.

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PHYLOGENETIC METHOD: Step 1

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PHYLOGENETIC METHOD: Step 2

For *n* diploid individuals, assume the following model for trait data, $Y_{n\times1}$:

$$
\begin{array}{rcl} \mathbf{Y} & = & \mathbf{Y_g} + \mathbf{Y_e} \\ \mathbf{Y_g} & \sim & N \left(Z D \mu, Z V Z^T \sigma^2 \right) \end{array}
$$

Phylogenetic Tree Parameters:

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$$

Phylogenetic Tree Parameters:

$$
D(\Theta) = 2n \times k \text{ matrix with elements:}
$$
\n
$$
D_{ij} = \begin{cases} 1, & \text{if tip } i \text{ is in cluster } j \\ 0, & \text{otherwise} \end{cases}
$$
\n
$$
\mu(\Theta) = (\mu_1, \mu_2, \dots, \mu_k)^T = \text{vector of within-cluster trait means}
$$
\n
$$
V(\Theta) = \text{variance-covariance structure determined by the estimated phylogeny,}
$$
\n
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PHYLOGENETIC METHOD: Parameter Example $(k = 3)$

$$
\mu(\Theta) = (\mu_1, \mu_2, \mu_3)^T
$$

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PHYLOGENETIC METHOD: Parameter Example $(k = 3)$

$$
\mu(\Theta) = (\mu_1, \mu_2, \mu_3)^T
$$
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$$
A \begin{bmatrix} j = 1 & j = 2 & j = 3 \\ 1 & 0 & 0 \\ B & 1 & 0 & 0 \\ D(\Theta) = C & 0 & 1 & 0 \\ D & 0 & 0 & 1 \\ E & 0 & 0 & 1 \\ F & 0 & 0 & 1 \end{bmatrix}
$$

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$$
\n
$$
F\begin{bmatrix}\nA & B & C & D & E & F \\
0 & 0 & 1 & 0 \\
0.29 & 0.41 & 0.06 & 0 & 0 \\
0.29 & 0.41 & 0.06 & 0 & 0 & 0 \\
E & 0.06 & 0.06 & 0.41 & 0 & 0 & 0 \\
0 & 0 & 0 & 0 & 0.41 & 0.07 & 0.07 \\
E & 0 & 0 & 0 & 0.07 & 0.07 & 0.41 \\
E & 0 & 0 & 0 & 0 & 0.07 & 0.07 & 0.41 \\
E & 0 & 0 & 0 & 0 & 0.07 & 0.041 & 0.07\n\end{bmatrix}
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$$

Genetic Component Notation/Parameters:

 $Z_{n\times2n}$ = $n\times2n$ matrix that maps each tip to diploid individual assuming each chromosome contributes equally to Y_{σ}

$$
\sigma^2 = \text{variance due to genetic component of trait}
$$

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$$

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$$
\mathbf{Y_g} \sim N(ZD\mu, ZVZ^T\sigma^2)
$$

\n
$$
\mathbf{Y_e} \sim N(X\beta, I\nu^2)
$$

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\mathbf{Y_e} & \sim & N\left(X\beta, l\nu^2\right)\n\end{array}
$$

Environmental Parameters: п.,

$$
\mathbf{X}^T = \begin{bmatrix} 1 & 1 & \dots & 1 \\ X_1 & X_2 & \dots & X_n \end{bmatrix}
$$

where X_i = value of the covariate for the *i*th observation

$$
\beta = (\beta_0, \beta_1)^T
$$
 = vector of regression coefficients

$$
\nu^2 = \text{variance due to environmental component of trait}
$$

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PHYLOGENETIC METHOD: Step 3

Estimation in a Bayesian Framework

The Likelihood:

$$
\mathbf{Y} = \mathbf{Y_g} + \mathbf{Y_e}
$$

where $\mathbf{Y} | \mathbf{Y_g}, \mu, \beta, \nu^2, \sigma^2 \sim N(\mathbf{X}\beta + \mathbf{Y_g}, \nu^2 \mathbf{I})$ and
 $\mathbf{Y_g} | \mu, \beta, \nu^2, \sigma^2 \sim N(\mathbf{ZD}\mu, \sigma^2 \mathbf{ZVZ}^T)$

Prior Distributions:

- $\beta \sim N(\beta_0, u^2 I)$ $\mu \sim N(\mu_0, w^2 I)$ $\sigma^2 \sim$ Inverse Gamma (*Shape = a, Scale = b*) $\nu^2 \sim$ Inverse Gamma $(\mathit{Shape}=c,\mathit{Scale}=d)$
- and we assume all parameters are independent!

Note: The **conditional posterior distributions** have closed forms!

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PHYLOGENETIC METHOD

Advantages of the phylogenetic method:

- Allow for and uses covariance among the observations
- Clustering uses the broad-scale evolutionary relationships to remain computationally feasible
- Bayesian framework produces posterior means for estimates

Ways to Assess Performance of Phylogenetic Method:

- **•** Simulation Study
- Real Data Analysis

METHODS: Data Simulation

Data needed for a simulation study include:

- **1.** SNP data
- 2. Covariate data
- **3.** Quantitative trait data that has
	-
	- **a.** a genetic component (related to a single SNP)
	- **b.** an environmental component (related to an external covariate)

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DATA SIMULATION METHOD

6. Simulate SNP data.

 $\left\{ \begin{array}{ccc} 1 & 0 & 0 \\ 0 & 1 & 0 \end{array} \right.$, $\left\{ \begin{array}{ccc} \frac{1}{2} & 0 & 0 \\ 0 & 0 & 0 \end{array} \right.$

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DATA SIMULATION METHOD

9. Simulate SNP data.

 \bullet Simulate covariate values (X) for each diploid individual uniformly from a specified range.

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$$
\begin{array}{rcl} \mathbf{Y} & = & \mathbf{Y_g} + \mathbf{Y_e} \\ & = & \rho \mathbf{T_g} + (1 - \rho) \mathbf{T_e}, \end{array}
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- \bullet \mathcal{T}_{g} : Genetic Component
	- Simulate data along the "disease" tree using a two-target Ornstein-Uhlenbeck process.

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9. Simulate SNP data.

- **2.** Simulate covariate values (X) for each diploid individual uniformly from a specified range.
- **3.** Simulate quantitative trait data: For some $\rho \in [0,1]$, let

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$$

- \bullet \mathcal{T}_{g} : Genetic Component
	- Simulate data along the "disease" tree using a two-target Ornstein-Uhlenbeck process.
	- Randomly pair the tips to create individuals.

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- \bullet \mathcal{T}_{g} : Genetic Component
	- Simulate data along the "disease" tree using a two-target Ornstein-Uhlenbeck process.
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	- Average the trait across SNP copies to find T_{α} .

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	- Randomly pair the tips to create individuals.
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\bullet \mathcal{T}_e : Environmental Component

 $T_e \sim N(X\eta, \tau^2 I)$, where η and τ^2 are fixed.

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SIMULATION STUDY: Data Simulation

Data Simulation Process:

- **1.** Simulate chromosomes in an ARG framework (SNP data).
- 2. Simulate covariate data
- **3.** Simulate quantitative trait data.
	- Simulate the genetic and environmental components of the trait.
	- Take a weighted average of these components to find the quantitative trait value.

Simulated Data:

- A matrix of SNP values at tips of phylogenies
- A vector of covariate values for each diploid individual.
- \bullet A vector of quantitative trait values, Y, for each diploid individual.

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DATA SIMULATION METHOD

In the interim steps, the data looks like this:

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DATA SIMULATION METHOD

The observed data is:

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SIMULATION PARAMETERS

Parameters for Genetic Component of Trait:

- $Y_i(0) = 90$
- $\delta_1 = 80, \delta_2 = 100$
- $\alpha = 10$
- $\sigma_{\rm V} = 20$

Parameters for Environmental Component of Trait:

- $r = 1$ covariate
- \bullet X_i are independent draws from a Uniform(25, 35) distribution
- $\boldsymbol{\eta}=(\eta_0,\eta_1)^{\textstyle \mathcal{T}}=(10,2.5)^{\textstyle \mathcal{T}}$
- $\bullet \tau = 15$
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SIMULATION STUDY: DATA ANALYSIS PROCESS

For each simulated data set:

- Using the phylogenetic tree, trait, and covariate data, estimate parameters using the posterior means from the Gibbs sampler.
- Note: True trees are used in this simulation study and the number of clusters is set to $k = 5$.

RESULTS: Known Phylogenies, Informative Priors

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RESULTS: Estimated Phylogenies, Informative Priors

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RESULTS: Known Phylogenies, Vague Priors

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RESULTS: Real Data Analysis

- Organisms: 91 wild-caught mice
- **Genetic Data: SNP data**
- Quantitative Traits: nine quantitative color phenotypes
- Covariates: Include weight, body length, tail length
- Goal: To identify regions of the genome contributing to mouse coat color after accounting for population structure covariates.
- **Previous work showed that much of** coat color variation appears to be controlled by a single gene, Agouti.

REAL DATA ANALYSIS ALGORITHM

For the real data set:

- **Computationally phase the data using Beagle.**
- At each SNP, estimate the phylogenetic tree using Blossoc and branch lengths using approximate MLEs.
- Using the phylogenetic tree, trait, and covariate data, estimate the parameters using the posterior means from the Gibbs sampler.
- Note: Estimated trees are used in this simulation study and the number of clusters is set to $k = 5$.

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RESULTS: Real Data Analysis

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RESULTS: Real Data Analysis

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CONCLUSIONS

- Posterior means provide good estimates of environmental parameters, even when two SNPs are considered.
- Using an evolutionary framework to approach problem is more realistic than non-tree based approximations.
- Use of the broad-scale evolutionary relationships among SNPs makes the technique computationally feasible.
- This model allows for analysis on a per-individual basis while preserving per-chromosomal estimation of evolutionary history at each SNP.

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FUTURE DIRECTIONS

Related problems of interest:

- the analysis of related genetic and environmental components
- the study of multivariate traits (multiple traits affected by one SNP)
- **•** developing a way to control for other associated SNPs present in the genetic data
- the analysis of data with population structure

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Thank You! Questions?

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- **Linnen, C. R. et al. 2013. Adaptive Evolution of Multiple** Traits Through Multiple Mutations at a Single Gene. Science, 339(6125):1312–6.
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Supplemental Results Modeling External Covariates

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DATA ANALYSIS ALGORITHM

LSS-C/LSS-I: At each SNP site, do the following.

- Estimate the marginal tree topology and branch lengths.
- Calculate LSS-C/LSS-I using the estimated phylogeny, covariate data, and trait data.

Previous Methods: At each SNP site, calculate the Likelihood

Ratio Test Statistic and the p-value from SNPassoc.

Data Analysis for Each Method:

- Detection of SNP/Covariate Analysis: Use permutation testing to check if any SNP along the chromosome or the covariate is detected.
	- Permute the trait values across the tips of the estimated phylogeny.
	- Recalculate each statistic using the permuted data.
- Localization Analysis: Record distance (in base pairs) between the most maximally-scored SNP and th[e a](#page-57-0)[ss](#page-59-0)[oc](#page-57-0)[ia](#page-58-0)[t](#page-59-0)[e](#page-53-0)[d](#page-54-0) [S](#page-72-0)[N](#page-53-0)[P](#page-54-0)[.](#page-72-0)

RESULTS: Detection and Localization

Figure: Power and localization in covariate analysis

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RESULTS: Example Replication

Figure: Example of behavior of LSS-C across a chromosome

Legend:

- Truly-associated SNP (located at red line) and related environmental covariate present
- No associated SNP nor related environment[al](#page-59-0) c[ov](#page-61-0)[a](#page-59-0)[ria](#page-60-0)[t](#page-61-0)[e](#page-53-0)[pre](#page-72-0)[s](#page-53-0)[e](#page-54-0)[nt](#page-72-0)

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RESULTS: Example Replication

Figure: Behavior of within-cluster mean estimates along a chromosome (using the chromosomal model)

- Truly-associated SNP and related covariate present
- **b** Neither a truly-associated SNP nor a related covariate present

RESULTS: Estimates at the Maximally-Scored SNP

Figure: Estimates of genetic and environmental variances at the maximally-scored SNP

(a) LSS-C

LSS-I

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RESULTS: Estimates at the Maximally-Scored SNP

Figure: Estimated differences in cluster means at the maximally-scored SNP LSS-C

LSS-I

RESULTS: Estimates at the Maximally-Scored SNP

[−] [−] [−] [−] 100 50 100 Figure: Estimates of $\beta_0 + \mu_k$ at the [−] [−] [−] [−] [−] ● maximally-scored SNP 50 LSS-C $\hat{\mathbb{B}}_{\mathrm{o}} + \hat{\mathbb{H}}_{\mathrm{e}}$ LSS-I −50 ρ=0 ρ=0.25 ρ=0.5 ρ=0.75 ρ=1 (a) − − − − − − − $\hat{\beta}_0 + \hat{\mu}_k$ 50 − − −100 $\rho = 0$ $\rho = 0.25$ $\rho = 0.5$ $\rho = 0.75$ $\rho = 1$ (b) つくへ 4日) 4. 点 Ξ 性 ∍

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RESULTS: Estimates at the Maximally-Scored SNP

Figure: Estimates of β_1 at the maximally-scored SNP

(a) LSS-C

(b) LSS-I

RESULTS: Example Replication

Figure: Example of number of clusters chosen by LSS across a chromosome

Legend:

- Truly-associated SNP (located at red line) and related environmental covariate present
- No associated SNP nor related environmental covariate present

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RESULTS: Example Replication

Figure: Example of behavior of baseline estimate across a chromosome.

Legend:

- Truly-associated SNP (located at red line) and related environmental covariate present
- No associated SNP nor related environment[al](#page-66-0) c[ov](#page-68-0)[a](#page-66-0)[ria](#page-67-0)[t](#page-68-0)[e](#page-53-0)[pre](#page-72-0)[s](#page-53-0)[e](#page-54-0)[nt](#page-72-0)

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RESULTS: Example Replication

Figure: Example of behavior of estimation of β_1 across a chromosome

Legend:

- Truly-associated SNP (located at red line) and related environmental covariate present
- No associated SNP nor related environmen[tal](#page-67-0) [co](#page-69-0)[v](#page-67-0)[ari](#page-68-0)[a](#page-69-0)[t](#page-53-0)[e](#page-54-0) [pr](#page-72-0)[e](#page-53-0)[s](#page-54-0)[en](#page-72-0)[t](#page-0-0)

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RESULTS: Adjusting for External Covariates

Table: Type I error of LSS and SNPassoc when adjusting for environmental covariates

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RESULTS: Adjusting for External Covariates

Figure: Power and localization when adjusting for covariates

Statistics: SNPassoc, LSS

RESULTS: Adjusting for External Covariates

Table: Power of Detection of LSS and SNPassoc when adjusting for environmental covariates

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RESULTS: Adjusting for External Covariates

Table: Average localization distance (bp) of LSS and SNPassoc when adjusting for environmental covariates

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